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2 **SCIENTIFIC WORKING GROUP ON DNA ANALYSIS**  
3 **METHODS<sup>1</sup>**

4 ***SWGDM Validation Guidelines for DNA Analysis***  
5 ***Methods: Overview Document***

6  
7 **Short Title: *Validation Overview Document***

8 **Effective XXXXXXX, XX, XXXX**

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10 **Scope**

11 The SWGDAM Validation Guidelines for DNA Analysis Methods: Overview Document  
12 provides guidelines for the validation of DNA analysis methods and supersedes the  
13 Scientific Working Group on DNA Analysis Methods (SWGDM) Validation  
14 Guidelines for Forensic DNA Analysis Methods (2016). These guidelines are intended to  
15 serve as instructions for laboratories in validating procedures consistent with the *FBI*  
16 *Director’s Quality Assurance Standards for Forensic DNA Testing and DNA Databasing*  
17 *Laboratories* (QAS). Each laboratory seeking to evaluate a new method shall determine  
18 which validation studies are relevant to the methodology, in the context of its application,  
19 and determine the experiments required to satisfy each study.

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<sup>1</sup> The Scientific Working Group on DNA Analysis (SWGDM; see [SWGDM.org](http://SWGDM.org)) is comprised of forensic science practitioners and other experts who represent government laboratories within the U.S and Canada, as well as intra- and international professional groups and academia. SWGDAM recommends to the FBI Director revisions to the *Quality Assurance Standards for Forensic DNA Testing Laboratories* and the *Quality Assurance Standards for DNA Databasing Laboratories* (QAS). SWGDAM provides a forum for its members and invited guests to discuss research, technologies, techniques, and training; and conduct or recommend studies to develop, test, and validate methods for use by forensic laboratories. SWGDAM’s Guidelines and Recommendations represent best practices within the discipline. The term “should” is used herein to indicate good practices identified by SWGDAM. “Shall” distinguishes mandatory elements, which may be specified in the Quality Assurance Standards for Forensic DNA Testing Laboratories and/or Quality Assurance Standards for DNA Databasing Laboratories.

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### Key Concepts:

- ❖ Each laboratory or laboratory system seeking to evaluate a new methodology shall determine which validation studies are relevant, in the context of its application, and determine the experiments required to satisfy each study.
- ❖ Validation shall precede the implementation of any new methods used for forensic DNA analysis.
- ❖ Developmental validation shall use case-type samples and include, as applicable, the following studies: characterization of genetic markers, species specificity, sensitivity, stability, precision and accuracy, population, mixture and PCR-based.
- ❖ Internal validation studies are used to supplement developmental validation and shall include the following studies, as applicable: known and non-probative evidence samples or mock evidence samples, sensitivity and stochastic, precision and accuracy, mixture and contamination.

### 1. Introduction

In the forensic context, the term “validation” refers to the process by which a procedure is evaluated to determine its efficacy and reliability for forensic application. This document

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58 and subsequent modules provide guidelines for the validation of DNA analysis methods  
59 and supersedes the Scientific Working Group on DNA Analysis Methods (SWGDM)  
60 Validation Guidelines for Forensic DNA Analysis Methods (2016). Terms used in this  
61 document and subsequent modules are intended to be consistent with definitions provided  
62 in the QAS.

63  
64 Because these are guidelines and not minimum standards, in the event of a conflict  
65 between the QAS and these guidelines, the QAS and the QAS Audit Documents have  
66 precedence. Additionally, to avoid any such conflict, the mandatory term ‘shall’ has been  
67 used when that term is similarly used in the QAS although the use of the term ‘shall’ is  
68 not intended to transform these guidelines into standards. Laboratories are encouraged to  
69 evaluate and update their standard operating procedures and validation approach as  
70 needed, in light of these guidelines.

71  
72 Methodology refers to the categories of methods used to perform a stage of DNA typing  
73 technology or technologies (e.g., methodologies for STR technology can include  
74 extraction, quantification, amplification, and detection,). Each laboratory seeking to  
75 evaluate a new method shall determine which validation studies are relevant to the  
76 methodology, in the context of its application, and determine the experiments required to  
77 satisfy each study. These guidelines are applicable to most methods used in DNA  
78 analysis. Some studies described herein may also assist in conducting evaluations of  
79 procedural modifications to existing validated methods.

80  
81 Performing internal validation studies can be a time consuming and laborious process.  
82 Laboratories are encouraged to communicate and discuss plans and experiences regarding  
83 validation workflows which may save time and resources.

84  
85 Laboratories validating new methods are encouraged to publish validation studies in a  
86 peer-reviewed journal or other means of dissemination to the forensic community.  
87 Publication provides access to information that other laboratories can use to guide their  
88 internal validation efforts. Utilization of published validation data from laboratories can  
89 increase efficiency, provide a valuable crosscheck between laboratories and enable  
90 ongoing improvements, and as a result, is strongly encouraged to promote consistency  
91 and demonstrate concordance among laboratories.

92  
93 These Validation Guidelines have been organized such that recommended elements of  
94 validation studies are contained herein (referred to as the “Overview” document). The  
95 Overview document will be supplemented by modules intended to provide technology or  
96 methodology specific guidance. These modules will be continually added or edited as  
97 necessary and will be posted to the SWGDAM website: [SWGDM.org/publications](http://SWGDM.org/publications). The  
98 studies in each module are not synchronized to the QAS; instead, they are presented in a  
99 suggested order to conserve resources such as time, reagents, samples and consumables  
100 and streamline required testing.

101  
102 The study examples provided in the module appendices are informational and are not  
103 intended to dictate the types and numbers of samples every laboratory must use to satisfy

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104 each study. Validation studies cannot account for all scenarios that may arise during  
105 casework examinations; however, laboratories should attempt to cover the range of  
106 variation expected to be encountered with forensic samples. Following implementation,  
107 laboratories should review results and, if necessary, conduct supplemental studies to  
108 improve workflow, analysis criteria, and/or interpretation.

## 109 110 111 **2. General Considerations**

112  
113 The purpose of validation is to demonstrate the reliability and potential limitations of a  
114 method. There are two types of validations required for method implementation for  
115 forensic DNA analysis – developmental and internal. The application of existing  
116 technology to the analysis of forensic samples does not necessarily create a new  
117 methodology. Published developmental validation studies in other fields may sufficiently  
118 address forensic applications.

119  
120 2.1 Developmental validation shall precede the implementation of any new methods  
121 used for forensic DNA analysis.

122  
123  
124 2.1.1 Peer-reviewed publication of developmental validation studies is strongly  
125 encouraged; however, validated methods may be implemented without  
126 such publication provided the underlying scientific principle(s) has been  
127 published.

128  
129 2.1.2 A DNA laboratory may rely upon another laboratory's published  
130 developmental validation studies. The citations and/or publications  
131 referencing that validation must be available and accessible to support the  
132 underlying scientific basis.

133  
134 2.2 Prior to using a method or procedure for forensic applications, a laboratory shall  
135 conduct internal validation studies on samples representative of those typically  
136 encountered by the end-user laboratory to demonstrate the reliability and potential  
137 limitations of the method.

138  
139  
140 2.2.1 Standard operating procedures, quality assurance parameters, guidelines  
141 for the evaluation and interpretation of analytical controls and DNA typing  
142 results, and as applicable statistical calculations, shall be derived from  
143 internal validation studies.

144  
145 2.2.1.1 For example, lower template DNA may cause extreme heterozygote  
146 imbalance; as such, empirical heterozygote peak-height ratio data  
147 could be used to formulate mixture interpretation guidelines and  
148 determine the appropriate ratio by which two peaks are determined to  
149 be heterozygotes.

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2.2.1.2 In addition to establishing an analytical threshold, results from sensitivity studies could be used to determine the extent and parameters of quality control tests that reagents or instruments require prior to their being used in actual casework.

152

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2.2.2 For laboratory systems that consist of more than one laboratory, each of the laboratories shall complete, document, and maintain studies which may be impacted by site-specific factors (e.g. precision, sensitivity, and contamination). Studies that are not location-specific may be shared among locations and the summary of the shared validation data shall be available at each site.

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2.2.3 It is important to utilize DNA samples extracted using the laboratory's validated methods as part of the internal validation studies.

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2.2.3.1 Control samples (e.g., HL60, 2800M, 9947A, SRM, 007, and others) are expected to behave differently than samples extracted using laboratory processes, therefore, the known samples included in a validation should not be exclusively control samples. Control samples can be used to supplement samples extracted using the laboratory's processes.

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### 3. Developmental Validation

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The developmental validation process shall include, where applicable, the following studies using samples that are representative of those typically encountered by the end user laboratory:

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178

179

**3.1 Characterization of genetic markers:** The basic characteristics (described below) of a genetic marker shall be determined and documented.

180

181

182

183

3.1.1 **Inheritance:** The mode of inheritance of DNA markers demonstrated through family studies.

184

185

186

3.1.2 **Mapping:** The genomic location of the genetic marker.

187

188

3.1.3 **Detection:** Technological basis for identifying the genetic marker (e.g., capillary electrophoresis, DNA sequencing, hybridization assays).

189

190

191

3.1.4 **Polymorphism:** Type of variation (e.g., sequence and/or length variants).

192

193

**3.2 Species specificity:** The ability to detect genetic information from non-human or non-targeted species (e.g., detection of microbial DNA in a human assay) shall be

194

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195 determined through laboratory studies and/or sequence homology searches against  
196 genomic databases (e.g., a BLAST search). The detection of genetic information  
197 from non-human or non-targeted species does not necessarily invalidate the use of  
198 the assay but may help define the limits of the assay.

199

200 **3.3 Sensitivity studies:** The ability to obtain reliable results from a range of DNA  
201 quantities, to include the upper and lower limits of the assay, shall be evaluated.

202

203 **3.4 Stability studies:** The ability to obtain results from DNA recovered from  
204 biological samples deposited on various substrates and subjected to various  
205 environmental and chemical insults should be evaluated. If substrates and/or  
206 environmental and/or chemical insults could potentially affect the method, then  
207 the method shall be evaluated to determine the effects of such factors.

208

209 3.4.1 For database samples, stability studies may include samples on various  
210 substrates and subjected to potential PCR inhibitors or various storage  
211 conditions.

212

213 **3.5 Precision and accuracy studies:** The ability of the assay to obtain repeatable  
214 and/or reproducible results must be determined, when practicable.

215

216 3.5.1 The measure of precision is usually expressed in terms of imprecision and  
217 computed as a standard deviation of the test results while the measure of  
218 accuracy can be accomplished by checking results against an appropriate  
219 and available certified reference material.

220

221 **3.6 Case-type samples:** The ability to obtain reliable results should be evaluated  
222 using samples that are representative of those typically encountered by the end-  
223 user laboratory. Where appropriate, consistency of typing results should be  
224 demonstrated by comparing results from the previous procedures to those  
225 obtained using the new procedure.

226

227 **3.7 Population studies:** The distribution of genetic markers in populations (i.e.,  
228 frequencies) must be determined in relevant population groups. Databases must  
229 be tested for independence expectations (e.g., Hardy Weinberg Equilibrium and  
230 Linkage Equilibrium).

231

232 **3.8 Mixture studies:** The ability to obtain reliable results from mixed-source samples  
233 shall be determined.

234

235 3.8.1 Studies should use mixture samples representing the number of  
236 contributors and the range of general mixture types expected to be  
237 encountered by the end-user laboratory.

238

239 3.8.1.1 These are best achieved by varying the number of contributors,  
mixture ratios, and overall template amounts.

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240 3.8.2 These studies will assist the laboratory in establishing guidelines for  
241 mixture interpretation, which may include estimation of the number of  
242 contributors, determination of the major and minor contributor profiles,  
243 and contributor ratios or proportions in addition to correlating  
244 male:autosomal or male:female DNA quantification determination with  
245 the expected STR results.  
246

#### 247 3.9 PCR-based studies:

248 PCR-based studies should include:

- 249
- 250 3.9.1 The reaction conditions needed to provide the required degree of  
251 specificity and robustness shall be determined. These include, but are not  
252 limited to, thermal cycling parameters, the concentration of primers,  
253 buffers, magnesium chloride, dNTPs and DNA polymerase.  
254
- 255 3.9.2 The potential for differential amplification among loci, preferential  
256 amplification of alleles within a locus, and stochastic amplification should  
257 be assessed to measure the specificity and robustness of the PCR reaction  
258 and the impact on peak height balance between and within a genetic  
259 marker.  
260
- 261 3.9.3 The effects of multiplexing should be assessed to measure the specificity  
262 and robustness of the PCR reaction.  
263
- 264 3.9.4 Appropriate controls should be assessed to ensure that the method works  
265 correctly and ensure the data are valid.  
266
- 267 3.9.5 Criteria for detection of amplified product should be determined based on  
268 the platform and/or method used and instrument baseline noise should be  
269 defined for quantitative and capillary electrophoresis typing methods.  
270
- 271 3.9.6 Appropriate measurement standards (qualitative and/or quantitative) for  
272 characterizing the alleles or resulting DNA product should be established.  
273
- 274 3.9.7 Publication of the sequence of individual primers is not required to  
275 appropriately demonstrate the reliability and limitations of PCR-based  
276 technologies. However, availability of the primer sequences is encouraged  
277 to aid in the identification of potential primer binding site variants and  
278 troubleshooting.  
279  
280

#### 281 4. Internal Validation

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283 The internal validation process shall include the applicable studies detailed below and  
284 outlined in the relevant module(s).

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**4.1 Known and non-probative evidence samples or mock evidence samples:**

- 4.1.1 Methods intended for casework samples shall be evaluated and tested using known samples (e.g., reference blood or buccal samples) and case-type samples. Mock evidence samples should be reflective of the range of types, quantity, and quality expected to be encountered in casework (e.g., various substrates, various concentrations, and degraded samples).
  - 4.1.1.1 Methods intended for database samples shall be evaluated and tested using known samples, available database samples, or mock samples collected on the substrates routinely encountered by the laboratory. Mock samples should be reflective of the types and quality expected to be encountered in databasing.
- 4.1.2 The known samples selected for the studies should exhibit a high level of heterozygosity. The use of heterozygous samples will help establish intra-locus balance metrics and aid in the determination of appropriate interpretation thresholds.
- 4.1.3 Known and non-probative sample studies may be used to:
  - assess the concordance of a method and therefore the degree of accuracy of the system.
  - help establish appropriate stutter filters
  - supplement the noise and threshold calculations
  - assess potential contamination events associated with the method
- 4.1.4 Case-type samples may include non-human DNA at template levels similar to those expected to be routinely encountered during casework analysis (e.g., mold, bacteria). Results of these studies can be used to determine how non-human artifacts can be recognized and how their presence will affect the interpretation of the DNA profile.
- 4.1.5 Results of these studies should be compared to previous results, where possible, to ensure concordance. Observed discordances should be documented, and where possible, an explanation should be provided.

**4.2 Sensitivity and Stochastic Studies:**

- 4.2.1 The laboratory shall determine the sensitivity levels of the assay or procedure.



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329 4.2.1.1 The known samples selected for the studies should exhibit a high level  
330 of heterozygosity. The use of heterozygous samples will help establish  
331 intra-locus balance metrics.

332  
333 4.2.1.2 Sensitivity studies can be used to:

- 334
- 335 • assess the ability to obtain reliable results from a range of
  - 336 DNA quantities, including the upper and lower limits of the
  - 337 assay
  - 338 • determine the dynamic range, ideal target range, limit of
  - 339 detection, heterozygote balance (e.g., peak height ratio),
  - 340 and the signal-to-noise ratio associated with the assay
  - 341 • evaluate excessive random (stochastic) effects generally
  - 342 resulting from low quantity and/or low-quality samples
  - 343

### 344 4.3 Precision and Accuracy Studies:

345  
346 4.3.1 Precision and accuracy of the assay/instrument shall demonstrate that it is  
347 generating the expected results. These studies should also address  
348 repeatability and/or reproducibility when practicable.

349  
350 4.3.1.1 **Repeatability:** Precision and accuracy of results (e.g., quantitative  
351 and/or qualitative) produced by the same operator and/or detection  
352 instrument should be evaluated.

353  
354 4.3.1.2 **Reproducibility:** Precision and accuracy of results (e.g., quantitative  
355 and/or qualitative) produced by different operators and/or detection  
356 instruments should be evaluated.

357  
358  
359 4.3.2 Precision depends only on the distribution of random errors and does not  
360 relate to the true value or specified value. The measure of precision is  
361 usually expressed in terms of imprecision and reported as the standard  
362 deviation of the test results.

363  
364 4.3.3 Accuracy of a measuring instrument is the ability of the instrument to give  
365 responses close to a true value. This can be accomplished by comparing  
366 the results against an appropriate and available certified reference  
367 material.

### 368 369 4.4 Mixture Studies:

370

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371 4.4.1 Mixture studies consisting of samples that are representative of those  
372 typically encountered by the laboratory shall be performed. For example,  
373 forensic DNA mixture studies should use samples that represent the  
374 number of contributors and the range of general mixture types for which  
375 the procedure will be used in casework (e.g., mixture proportions and  
376 template quantities).

377  
378 4.4.1.1 These studies must be used to establish interpretation guidelines to  
379 include estimation of the number of contributors to the mixture,  
380 determination of the major and minor contributor profiles, when  
381 appropriate, and for instituting criteria to deduce potential contributors.  
382

383 4.4.1.2 As an additional example, laboratories validating a new extraction  
384 method should include in the mixture studies the body fluids, and  
385 combinations thereof, that they typically encounter.  
386

#### 387 4.5 Contamination Assessment:

388  
389 4.5.1 Contamination studies shall be performed to evaluate and measure the  
390 potential for the introduction of exogenous DNA at any point during  
391 sample processing. Based on these studies, the laboratory should  
392 determine quality control procedures to mitigate contamination and/or  
393 develop a policy for data interpretation when contamination has been  
394 identified.  
395

396 4.5.2 These studies also serve to assess the presence of potential contaminants  
397 in the reagents used throughout the various sample processes in the  
398 laboratory as well as the efficacy of personal protective equipment and  
399 cleaning protocols.  
400

401 4.5.2.1 The laboratory shall evaluate, using negative controls and known  
402 samples, the detection of exogenous DNA originating from reagents,  
403 consumables, other samples, operator(s) and/or the laboratory  
404 environment.  
405

406 4.5.3 Should contamination be encountered, the origin of the event must be  
407 explored and should be characterized when possible.  
408

409 4.5.3.1 The validation should establish procedures that will minimize the  
410 occurrence of contamination events. Standard operating procedures  
411 should detail how to address contamination should it occur in  
412 casework analyses.  
413

414 4.6 If conducted within the same laboratory, developmental validation studies may  
415 satisfy some elements of the internal validation. In these cases, a laboratory's

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416 internal validation can be used to supplement any elements in which the  
417 developmental validation is insufficient.

418

419 4.7 The laboratory should evaluate the suitability of each study based on the  
420 methodology and/or application. If the laboratory determines that a study is not  
421 applicable, the reason(s) shall be documented in the validation summary. Using  
422 the specific module(s) as guidance, the laboratory should determine the  
423 appropriate number of samples, and the types of samples required for each study  
424 to demonstrate the potential limitations and reliability of the method.

425

426 4.7.1 A validation study cannot account for all potential casework scenarios;  
427 however, samples representing the range of forensic sample types  
428 expected to be routinely encountered by the laboratory should be selected  
429 for evaluation.

430

431 4.8 At the time of validating new DNA methods (from amplification through  
432 characterization), typing test kit, or platform instrument model, the laboratory  
433 shall check results from the new method/kit/platform for concordance with an  
434 appropriate and available certified reference material (or sample made traceable to  
435 the certified reference material) prior to the implementation of the method for  
436 forensic analysis.

437

438 4.9 Internal validation data may be shared by all locations in a multi-laboratory  
439 system. The summary of the shared validation data shall be available at each site.  
440 At a minimum, each laboratory in a multi-laboratory system shall complete,  
441 document, and maintain applicable site-specific precision and accuracy,  
442 sensitivity and stochastic, and contamination assessment studies.

443

444

445 4.10 Internal validation studies shall be documented and summarized. Internal  
446 validation studies shall be reviewed by the technical leader and the approval  
447 documented prior to implementing a procedure for forensic applications.  
448 Documentation, at a minimum, should include:

449

450 4.10.1 Summary of each study conducted.

451

452 4.10.2 Results of each study, including generated data.

453

454 4.10.3 Approval of the technical leader for implementation.

455

456

## 457 5. Procedure Modification

458

459 Procedure modification is an alteration of an existing and previously validated analytical  
460 procedure that may have a consequential effect(s) on analytical results. Examples of a

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461 procedure modification include: a decrease in reaction volume of an amplification kit or  
462 an increase in injection time for a genetic analyzer.

463

464 5.1. A procedure modification must be evaluated prior to use with forensic samples.  
465 The modified procedure must be evaluated by comparing it to the original  
466 procedure using similar samples to ensure concordance and ascertain the  
467 potential benefits.

468

469 5.2 The laboratory should define the appropriate sample number, sample type, and  
470 the studies necessary to evaluate the modification. The evaluation shall be  
471 documented, reviewed by the technical leader and the approval documented prior  
472 to implementation.

473

474 5.2.1 If the procedure modification is determined to have an impact on the  
475 efficacy or reliability of the forensic analysis (such as modifications that  
476 impact the efficacy of the PCR process or the detection of DNA types),  
477 additional internal validation studies (such as sensitivity and stochastic  
478 studies) may be necessary to demonstrate the continued reliability and  
479 potential limitations of the method.

480

481

## 482 **6. Performance Check**

483

484 A performance check is a quality assurance measure to assess the functionality of  
485 laboratory critical equipment and instruments that affect the accuracy and/or  
486 validity of forensic sample analysis.

487

488 6.1 A laboratory shall have and follow a documented program for conducting  
489 performance checks of critical instruments and equipment.

490

491 6.1.1 This program will document the laboratory protocol, the performance  
492 characteristics and acceptance limits.

493

494 6.1.2 The laboratory should evaluate the appropriate sample number and type to  
495 demonstrate the reliability of the instrument or equipment.

496

497 6.1.3 If the laboratory determines that a performance check study is not  
498 necessary, the justification should be documented.

499

500 6.1.4 A laboratory's evaluation may also determine that additional performance  
501 check studies are necessary due to unacceptable data.

502

503 6.1.5 The completion and subsequent approval/rejection of the performance  
504 check must be documented.

505

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- 506 6.2 At a minimum, critical instruments or equipment shall require annual  
507 performance checks.  
508
- 509 6.3 If service is performed on a critical instrument or equipment, a performance check  
510 is required before returning it to use for forensic analysis.  
511
- 512 6.4 If the physical location or the environment of the instrument has been changed  
513 (e.g., instrument moved to another room, significant remodeling of the room), a  
514 performance check should be completed before returning it to use for casework  
515 analysis.  
516
- 517 6.5 After an internal validation has been performed on a critical instrument, each  
518 additional critical instrument of the same make and model shall require, at a  
519 minimum, a performance check.  
520
- 521 6.5.1 The performance check should demonstrate that results are reproducible  
522 on the new critical instrument and that testing results associated with new  
523 critical instrumentation are comparable to testing results generated during  
524 the internal validation and acceptable for use within the laboratory.  
525
- 526 6.5.2 If the laboratory determines that the new critical instrument is not within  
527 acceptable parameters, then the laboratory must address the instrument  
528 and/or procedure to minimize or mitigate the difference.  
529

## 531 7. Software

532

- 533 7.1 Software or software tools used in a forensic laboratory that may have an impact  
534 on the analytical process, interpretation, or statistical calculations shall be  
535 validated to ensure the software fulfills its intended purpose and is suitable for use  
536 in the laboratory. This includes software used as a component of instrumentation,  
537 software used for the analysis and/or interpretation of DNA data, software used  
538 for statistical calculations and software tools (e.g., macros, workbooks, LIMS)  
539 used for analytical workflows. Additional functions and/or features of software  
540 not intended for use by the laboratory do not require validation.  
541
- 542 7.1.1 Software shall be evaluated to assess its suitability for its intended use in  
543 the laboratory and to determine the necessity of validation studies and/or  
544 software testing. This evaluation shall be documented to include the  
545 determination of which studies will be conducted.  
546
- 547 7.1.2 Developmental validation shall be required for any software or new  
548 software modules used as a component of instrumentation, for the analysis  
549 and/or interpretation of DNA data, or for statistical calculations prior to  
550 implementation. At a minimum, the validation must include functional

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551 and reliability testing, and as applicable, accuracy, precision, sensitivity,  
552 and specificity studies.

553

554 7.1.3 Internal validation studies may include:

555

556 7.1.3.1 **Functional testing** to confirm that a software performs the tasks as  
557 expected.

558

559 7.1.3.2 **Reliability testing** to establish that the software can run in the  
560 laboratory's environment.

561

562 7.1.3.3 **Accuracy and precision studies** to ensure the software is making  
563 accurate measurements and/or correct calculations.

564

565 7.1.3.4 **Sensitivity studies** to evaluate the upper and lower limits of the  
566 software.

567

568 7.1.3.5 **Specificity studies** to evaluate the ability of the system to provide  
569 reliable results over a broad variety of typing results.

570

571 7.1.4 Software validations including the summary and results shall be reviewed  
572 by the laboratory's technical leader and approval documented prior to  
573 implementation.

574

575 7.2 Modifications to software, or a software upgrade, used as a component of  
576 instrumentation, for the analysis and/or interpretation of DNA data, or statistical  
577 calculations shall be evaluated to determine if the modifications result in major  
578 or minor revisions to the software. For software upgrades or modifications, the  
579 laboratory should require a software developer to provide written  
580 documentation, such as release notes, to explain the purpose and scope of the  
581 modification.

582

583 7.2.1 The requirement for validation and/or software testing is determined by  
584 the type of software change and the impact of the change on the operation  
585 of the software.

586

587 7.2.1.1 A *major* revision to software or software tools that are used as a  
588 component of instrumentation, for the analysis and/or interpretation of  
589 DNA data, or statistical calculations shall require validation prior to  
590 implementation. These validation studies shall include functional  
591 testing, reliability testing, regression testing, and, as applicable,  
592 precision and accuracy, sensitivity and specificity studies.

593

594 7.2.1.2 A *minor* revision to software or software tools that does not impact the  
595 analytical process, interpretation, or statistical calculations shall require

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596 at a minimum, a functional test prior to implementation to confirm that  
597 the software performs the tasks as expected.

598  
599 7.2.1.2.1 Operating system or security patches that are compatible with  
600 the system requirements of the software do not fall into the  
601 scope of these guidelines.

602  
603 7.3 Software validation studies may be shared by all locations in a multi-laboratory  
604 system. The summary of the shared validation data shall be available at each  
605 site. At a minimum, each laboratory in a multi-laboratory system shall  
606 complete, document, and maintain applicable site-specific reliability testing.  
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## Appendix A

### References and Suggested Readings

Butler, J.M., (2011) *Quality Assurance and Validation*. Advanced Topics in Forensic DNA Typing: Methodology. Elsevier.

Federal Bureau of Investigation. (2020) *Quality Assurance Standards for Forensic DNA Testing Laboratories*; available at <https://www.fbi.gov/about-us/lab/codis/qas-standards-for-forensic-dna-testing-laboratories-effective-07-01-2020>.

Federal Bureau of Investigation. (2020) *Quality Assurance Standards for Forensic DNA Databasing Laboratories*; available at <https://ucr.fbi.gov/lab/biometric-analysis/codis/quality-assurance-standards-for-forensic-dna-testing-laboratories>

Scientific Working Group on DNA Analysis Methods. (2017) *SWGDM Interpretation Guidelines for Autosomal STR Typing by Forensic DNA Testing Laboratories*; available at [https://1ecb9588-ea6f-4feb-971a-73265dbf079c.filesusr.com/ugd/4344b0\\_50e2749756a242528e6285a5bb478f4c.pdf](https://1ecb9588-ea6f-4feb-971a-73265dbf079c.filesusr.com/ugd/4344b0_50e2749756a242528e6285a5bb478f4c.pdf).

**Informational Web Site:** Additional information may be obtained from the following web site: <https://strbase.nist.gov/>



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## Appendix B

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### SWGDM Internal Validation Guideline Modules

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The Validation Guidelines for DNA Analysis Methods have been organized such that recommended elements of validation studies are contained in the “Overview” document. This Overview document is supplemented by modules intended to provide technology or methodology specific guidance. The study examples in each module are not synchronized to the FBI QAS nor are they intended to be prescriptive. Instead, they are presented in a suggested order to conserve resources such as time, reagents, samples, and consumables and to streamline required testing.

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#### **Internal Validation Module for an Autosomal Multiplex Kit (xxxx, 2025)**

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This module describes the recommended studies for validating an autosomal multiplex amplification/typing kit. Study purpose, considerations, examples, and outcomes are presented in a suggested order.

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#### **Internal Validation Module for a Fully Continuous Probabilistic Genotyping Systems (xxx, 2025)**

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This module describes the recommended studies for validating the use of fully continuous probabilistic genotyping systems (PGS) for analyzing DNA single source and mixture profiles by inferring genotype weights using algorithms and assigning likelihood ratios (LR(s)) to the comparison of known reference samples to a forensic sample. Study purpose, design/considerations and outcomes are presented in a suggested order.

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#### **Internal Validation Module for Quantitation Module (in progress)**

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#### **Internal Validation Module for Modified Rapid DNA for Analysis of Database, Known or Casework Reference Samples (in progress)**

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#### **Internal Validation Module for Next Generation Sequencing (in progress)**

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Document Version	Revision History
July 2003	Original. (Published in Forensic Science Communications in July 2004; available at <a href="http://www.fbi.gov/about-us/lab/forensic-science-communications/fsc/july2004/index.htm/standards/2004_03_standards02.htm">http://www.fbi.gov/about-us/lab/forensic-science-communications/fsc/july2004/index.htm/standards/2004_03_standards02.htm</a> )
November 2012	The document was revised to update the guidelines to incorporate changes to the FBI Director's Quality Assurance Standards (QAS). The revisions include: addition of a preface that describes the QAS have precedence over these guidelines; definitions added to Section 1 for critical instrument, methodology, precision and technology; revised description of developmental and internal validation in Section 2; added Table of recommended studies for internal validation in Section 4; and References and Suggested Reading added in a new Section 8.
November 2012	Approved by the SWGDAM membership.
December 2012	Approved by the SWGDAM Executive Board, with minor revisions, for posting on <a href="http://swgdam.org">swgdam.org</a> .
November 2016	The document was revised to address Next Generation Sequencing (NGS) technologies. Revisions include: new definitions in Section 1 for bioinformatics, index, library and next generation sequencing; revisions to the definitions in Section 1 for methodology and technology; the addition of NGS-specific studies to both Sections 3 and 4; and revisions to Section 7.
December 2016	Approved by the SWGDAM Executive Board, with minor revisions, for posting on <a href="http://www.swgdam.org">www.swgdam.org</a> .
December 2024	This document was reformatted to a Validation Overview document with general information about validation testing. The glossary was removed. References and Suggested Readings – formerly section 8 – has become the new Appendix A. Specific technology or methodology validation information has been moved to a separate Module format for each topic, reflected in the new Appendix B.

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